



Plant Archives

Journal homepage: <http://www.plantarchives.org>

DOI Url : <https://doi.org/10.51470/PLANTARCHIVES.2025.v25.supplement-2.136>

EFFECT OF NURSERY GROWING MEDIA FOR QUALITY SEEDLING PRODUCTION OF MARIGOLD, CALENDULA, DIANTHUS, AND SALVIA

Dweepjyoti Sarkar¹, Preeti Hatibarua², Ruchika Borbaruah^{3*}, Madhumita Choudhury Talukdar², Dharmendra Naikwad⁴ and Dhruva Jyoti Nath⁵

¹Agricultural Development Officer, Department of Agriculture, Majuli, Government of Assam-785106, India

²Department of Horticulture, Assam Agricultural University, Jorhat, Assam-785013

³Department of Floriculture and Landscaping, Bidhan Chandra Krishi Viswavidyalaya, Nadia, West Bengal-741252, India

⁴Section Officer Horticulture, Horticulture sub division, Central PWD, Ministry of Urban Development, Government of India

⁵Department of Plant Pathology, Assam Agricultural University, Jorhat, Assam-785013, India

*Corresponding author E-mail: ruchikab2018@gmail.com

(Date of Receiving : 17-03-2025; Date of Acceptance : 28-05-2025)

ABSTRACT

The expanding ornamental nursery industry relies on the growth medium as a critical factor influencing seed germination, seedling emergence, development, and quality. This document examines appropriate nursery growth mediums for the production of healthy and disease-free seedlings of four ornamental plant species viz. Marigold cv. Inca Orange, Calendula cv. Bon Bon Mix, Dianthus cv. Idea Select Mix, and Salvia cv. Vista Mix. The experimental treatments comprised four distinct media blends utilized in plastic plug trays maintained under greenhouse conditions in Assam, namely: M₁: Cocopeat (60): Vermiculite (20): Perlite (20), M₂: Cocopeat (60): Vermicompost (40), M₃: Cocopeat (60): Vermicompost (40): Microbial consortia (10 g/kg media), M₄: Traditional nursery (Soil: Sand: FYM in a 1:1:1 v/v ratio). Among the four-nursery medium M₃, there was a notable increase in seedling production characterized by earlier seedling emergence (3.16, 3.18, 4.10, and 2.84 DAS for marigold, calendula, dianthus, and salvia), seedling height, a greater number of leaves per seedling, as well as an increased quantity of roots per seedling, extended root length, and elevated fresh and dry root weight (0.42, 0.38, 0.38, 0.46 g fresh weight and 0.13, 0.14, 0.12, 0.15 g dry weight). This ultimately resulted in enhanced seedling fresh and dry weight (1.57 g, 1.57 g, 1.64 g, 1.31 g fresh weight and 0.40 g, 0.38 g, 0.37 g, 0.34 g dry weight), alongside increased chlorophyll content (1.57, 1.45, 1.54, and 1.56 mg g⁻¹ fresh weight), and ultimately resulted in improved in seedling vigor in M₃.

Keywords : nursery media, marigold, calendula, dianthus, salvia, microbial consortia

Introduction

Floriculture represents an escalating agro-industry in India. Its floriculture market is currently experiencing a compounded annual growth rate (CAGR) of 20.1% from 2019 to 2024 (Kumar *et al.*, 2024). The availability of diverse and conducive agro-climatic conditions, inexpensive and abundant labor, a well-established knowledge base, vast research network and traditions makes India in advantageous position to produce flowers year-round. Among the frequently cultivated flowers in India, the most

significant include marigold, aster, chrysanthemum, rose, carnation, jasmine, crossandra, tuberose, gladiolus, orchid, and others, which are extensively cultivated in states such as Tamil Nadu, Karnataka, Andhra Pradesh, West Bengal, Bihar, Rajasthan, Delhi, Uttar Pradesh, and Maharashtra (Thakur *et al.*, 2023). Within this category of ornamental plants, annual species such as marigold, pot marigold, dianthus, and salvia, which thrive in subtropical climates, are well-suited for farming in Assam. Nevertheless, the nursery

production in the state has not yet reached a standardized level.

Marigold (*Tagetes minuta* L.) is a highly favored annual flowering plant in Assam, flourishing in warm environments with full sunlight and exhibiting moderate resilience to drought once it has been established. It is classified within the Asteraceae family and commercially propagated via seeds. It is extensively cultivated as an ornamental flower, loose or cut flower, bedding, pot or landscape plant. The visually appealing and vividly colored flowers are utilized in the creation of garlands, religious offerings, exhibitions, and various decorations. Its petals are edible and serve as a source of commercial lutein (Sharma *et al.*, 2024; Manzoor *et al.*, 2022). In addition, chemical compounds known as 'thiopenes' extracted from roots of *Tagetes minuta* L. function as mosquito repellents (Walia and Kumar, 2020), while alpha-terthienyl derived from its root exhibits nematocidal, insecticidal, antiviral, and cytotoxic properties (Sachin and Homraj, 2021). Consequently, the integration of marigold as an intercrop and trap crop alongside species such as Coconut (Nath *et al.*, 2019), tomato (Kalaiselvam and Devaraj, 2011), and white cabbage (Jankowska *et al.*, 2009) are proved advantageous and in trend. Another notable annual plant in the region is Calendula (*Calendula officinalis*), commonly referred to as pot marigold, belongs to the Asteraceae family. This species is native to Southern Europe and is also propagated commercially via seeds. Calendula is cultivated in garden beds, as potted plants, as cut flowers, and in window boxes. The alcoholic extract from its leaves and flowers possesses antimicrobial properties and is employed in the treatment of varicose ulcers and skin lesions (Ullah *et al.*, 2023). Furthermore, the petals are edible and exhibit anti-helminthic and anticancer attributes (Pires *et al.*, 2018). One additional widely recognized winter annual is Sweet William (*Dianthus barbatus* L.), which is a triploid (3x) species exhibiting various forms and colors, classified within the family Caryophyllaceae. It is commercially cultivated through seeds and is commonly utilized as a bedding plant for landscaping purposes, as well as for pot plants and cut flower production. This species serves as an alternative to *Dianthus chinensis* in the treatment of menostasis, gonorrhea, cough, and has emmenagogue and diuretic properties (Xing *et al.*, 2015). Another prevalent annual ornamental from the Lamiaceae family is *Salvia splendens*. This plant is also propagated by seeds and has a long-standing application in traditional medicine for combating fever, rheumatism, perspiration, sexual dysfunction, and chronic bronchitis or various mental disorders (Miraj and Kiani, 2016). It is regarded as an

important ornamental crop for landscaping, with some varieties being utilized for the extraction of diverse culinary aromas. These plants exhibit drought resistance and flourish in sunny environments, making them suitable for the warm climate of Assam.

With the escalating demand for ornamental plants in both urban and rural regions of India, the pressure on nurseries has intensified. However, the production of flower seedlings using traditional methods induces stress in the plants. To address the constraints associated with soil-based production systems (such as soil-borne pests and other chemical and biological variabilities), the utilization of soilless growing media can significantly enhance seedling production under controlled conditions. Growing media is a critical factor that affects seed germination, seedling emergence, growth, and overall quality of seedlings in a nursery (Mathowa *et al.*, 2016). The quality of seedlings produced in a nursery impacts their re-establishment in the field and the subsequent productivity of an orchard (Mahmoud *et al.*, 2019). Growing media can comprise organic substances such as peat, compost, tree bark, coconut (*Cocos nucifera* L.) coir, and poultry feathers, or inorganic substances such as clay, perlite, vermiculite, and mineral wool. In addition to growing media, a revolutionized use of microbial consortia has emerged as a trend in nursery production. This primarily encompasses nitrogen-fixing bacteria such as *Azotobacter*, *Bacillus*, *Clostridium*, and *Klebsiella*; phosphate-solubilizing bacteria such as *Azospirillum*, *Bacillus*, *Pseudomonas*, *Rhizobium*, alongside newly identified endophytes and other microorganisms. Its integration into nursery practices for bedding ornamental annuals may lead to enhanced seed germination, improved root growth, increased efficiency of nutrient absorption, suppression of diseases, greater resilience to stress, and a decreased dependence on chemical inputs, as evidenced by numerous experimental investigations conducted on various other crops and plants (Rasool *et al.*, 2024; Mellidou and Karamanoli, 2022; Abdul *et al.*, 2007). These benefits play a significant role in producing robust, healthy, and high-quality bedding plants that are ready for transplantation and successful establishment in diverse landscapes or gardens.

The importance of quality ornamental seedling production cannot be over-emphasized. Therefore, standardization of processes for large-scale seed production, improving the germination of seeds, and other resources could help to overcome the prohibitive cost of these seedlings within the state of Assam. Therefore, a polyhouse experiment was conducted to determine the optimal growing media supplemented

with pre-sowing microbial consortia to yield quality and robust seedlings on portrays.

Materials and Methods

The current study was undertaken in the Experimental Farm, Dept of Horticulture, Assam Agricultural University, Jorhat (26.47°N latitude and 94.12°E longitude; 86.8 m above msl). The aim of this study was to standardize the most suitable nursery media for Marigold (Inca Orange), Calendula (Bon Bon Mix), Dianthus (Ideal Select Mix) and Salvia (Vista Mix). The experiment was carried out as Randomized Block Design, comprising four-nursery media composition i.e., Cocopeat, Vermiculite and Perlite at 60:20:20 ratio (M₁); Cocopeat and Vermicompost at 60:40 ratio (M₂); Cocopeat and Vermicompost at 60:40 ratio with Microbial consortia of *Azotobacter*, *Azospirillum* and Phosphate Solubilizing Bacteria (PSB) (10 g/kg media) (M₃); and Conventional nursery (Soil: Sand: FYM at 1:1:1 v/v) (M₄). One seed of each four ornamental in per cell of the pro-trays under protected conditions in the second week of October 2018.

Physico-chemical estimations of growing media

Various physico-chemical estimations of growing media performed during the experiment are as follows:

pH: The pH of media-water suspension was determined by systronic pH meter using combined glass and calomel electrode. Soil (20g) was mixed with 410 mL distilled water in 1:2 ratio and moved intermittently for 30 minutes. After this suspension was left undisturbed for 1 hr. In supernatant the combination electrode was put and pH was recorded.

Water holding capacity: Maximum water holding capacity of the soil was determined by using Keen's Raczowaski brass cup as described by Black, 1965.

Electrical conductance (EC): EC of soil-water suspension was determined by systronic conductivity meter. Soil sample (20g) was mixed in 40 mL distilled water. The suspension was stirred intermittently for 1/2hr and kept for 30minutes without any disturbance for complete dissolution of soluble salt. The soil was allowed to settled then conductivity cell inserted in solution to record the EC value.

CEC: Cation exchange capacity was determined by neutral normal ammonium acetate saturation method (Black, 1965). Down weight of soil was treated with neutral normal ammonium acetate solution which saturated the soil surface with ammonium ions and the excess ammonium ions present on non-exchangeable sites were washed with alcohol. Then the soil was

subjected to distillation and based on ammonia released CEC was computed.

Porosity %: Total porosity of soil was determined by using the values of particle density and bulk density (Black, 1965)

Seedling characteristics: The characteristics monitored during the experiment are Seedling emergence (%), Days to 2-True leaf stage, Days to transplanting, Seedling height (cm), Leaf area (cm²), Leaf number per plant, Leaf area index (LAI) by Evans, 1972, Root length (cm), root fresh weight (g), Root dry weight (g), Number of roots, Seedling fresh weight (g), Seedling dry weight (g) and Seedling growth index / seedling vigour index (SVI) as suggested by Abdul Baki and Anderson (1973).

Chlorophyll content (mg/g fw): Chlorophyll estimation was done from fresh leaf samples. The absorbance of the extract was measured at 645 and 663 nm wavelength filters in UVV spectrophotometer for the determination of total chlorophyll content according to the method developed by Singh and Singh and Singh (1977) as below:

Total chlorophyll in mg/gm tissue

$$= \frac{20.2 \times (D.645) + 8.02 \times (D.663) \times V}{1000 \times W}$$

Where,

V = Final volume of extract (mL)

W = Weight of sample taken (g)

D.645 = O.D. at 645

D.663 = O.D. at 663

Statistical analysis

The experiment was carried out in completely randomized design (CRD) at 5% level of significance with three replications. IBM-SPSS (www.spss.com.hk/software/statistics/data analysis) software was used to analyses the recorded data.

Results

Marigold

Different nursery growing media had considerable effect on seedling emergence and subsequent growth in Inca marigold. Among the four treatments, the optimal seedling attributes were noted in M₃ [Cocopeat (60): Vermicompost (40): Microbial consortia (10 g/kg media)]. Marigold seeds planted in trays within greenhouse conditions in M₃ experienced the shortest duration for emergence (3. 16 days), the least time for the formation of 2 true leaves and 4 true leaves (11. 50

and 28.81 days), indicating the fewest days required to achieve a transplantable stage during both the first and second years, as well as in the combined data. The cumulative data for marigold seedlings demonstrated (Table 1. and Table 2.) the highest seedling height (8.65 m), increased leaf area (6.29 cm²), leaf production (4.27 leaves/seedling), root production (38.23 roots/seedling), root length (9.39 cm), along with the highest fresh and dry weight of roots (0.42 and 0.13 g) and maximum seedling fresh weight (1.57 g) when cultivated in M₃. Consequently, this led to a heightened seedling vigor index of 2267.54 in M₃. Nonetheless, no notable difference in the turgidity values of seedlings grown in any substrate within greenhouse trays was observed under the conditions of Assam. Biochemically, a significantly higher chlorophyll content (1.57 mg g⁻¹ FW) was found in seedlings grown in M₃ (Fig. 1).

Calendula

In Calendula (Bon Bon Mix), during the years 2018-19 and 2019-20, as well as in the combined data, the optimal seedling performance was recorded in the treatment M₃ [Cocopeat (60): Vermicompost (40): Microbial consortia (10 g/kg media)]. Seeds germinated in the M₃ treatment exhibited the quickest emergence (3.18 days), resulting in the least time taken (11.47 and 27.63 days) for the formation of two leaves and progressing to the transplanting phase, specifically to the four-leaf stage (Table 1.). Furthermore, calendula seedlings cultivated in M₃ achieved the highest measurements in seedling height (9.34 cm), leaf area (7.12 cm²), leaf count (5.06 leaves/plant), leaf area index (1.39), root length (8.96 cm), and number of roots per seedling (33.97). This particular combination of cocopeat, vermicompost, and microbial consortia effectively nurtured the seedlings, leading to maximum root fresh and dry weights of 0.38 and 0.14 g, respectively, along with the greatest seedling fresh and dry weights of 1.57 and 0.38 g (Table 2.), respectively, which resulted in the highest seedling vigor index of 2221.35. However, no significant differences were observed in the turgidity values. Biochemically, the M₃ treatment also demonstrated a significantly elevated chlorophyll content of 1.45 mg/g FW (Fig 1.).

Dianthus

Seeds of Dianthus (Ideal Select Mix) sown in media M₃, M₂ and M₄ exhibited the earliest seedling emergence (4.10, 4.37 and 4.50 days in the pooled data) within the polyhouse conditions in trays, and these results were at par. Additional seedling traits like the least duration for the formation of 2 true leaves (9.71 days), the formation of 4 true leaves or the transplantable stage (28.21 days), the tallest seedling height (8.43 cm), leaf area (5.61 cm²), leaf area index (1.22), root length (9.12 cm), and production of leaves and roots (8.28 leaves per seedling and 39.06 roots per seedling), all enhanced by nursery media M₃. This media also achieved the highest root fresh and dry weight (0.38 and 0.12 g) and seedling fresh and dry weights (1.64 and 0.37 g), leading to a peak turgidity of 79.90% and an exceptional seedling vigor index of 2184.28 (Table 2.). Furthermore, plants cultivated in media M₃ achieved the significantly highest chlorophyll content (1.54 mg g⁻¹ FW in the pooled data analysis) (Fig. 1).

Salvia

Among the four types of sowing media, the best seedling performance of Salvia-Vista Mix cultivar were observed in M₃ media, which consists of Cocopeat (60%), Vermicompost (40%), and Microbial consortia (10 g/kg media). Salvia seeds planted in trays under greenhouse conditions within M₃ demonstrated the shortest duration for emergence (2.84 days), the quickest development of two true leaves (9.12 days), and attained the transplantable stage in the least time (28.16 days) during both the first and second years, as well as the cumulative data (Table 1.). Additionally, this media effectively supported seedlings, resulting in the highest recorded seedling height (6.22 cm), leaf area (5.85 cm²), leaf production (6.95 leaves/seedling), leaf area index (1.08), root length (9.44 cm), root production (65.38 roots/seedling), and ultimately, the highest root and seedling fresh and dry weights (Table 2.). Nonetheless, no significant differences were noted in the turgidity values of the seedlings. Biochemically, leaves of salvia grown in M₃ exhibited the significantly highest chlorophyll content (1.58 mg g⁻¹FW). Furthermore, the highest seedling vigor index of 2239.91 was recorded in M₃ (Fig. 1).

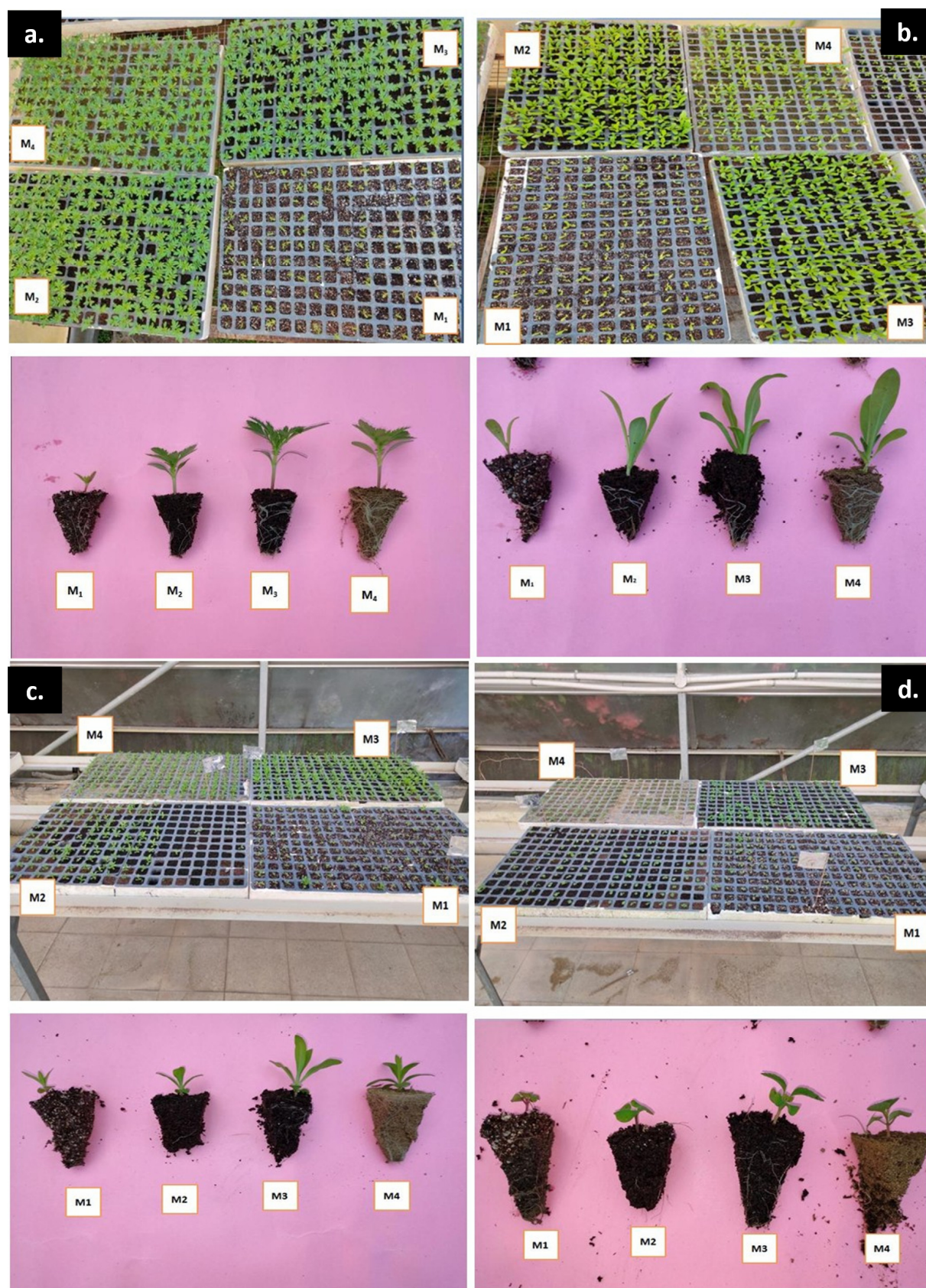


Fig. 1 : Seedlings of a. Marigold, b. Calendula, c. Dianthus and d. Salvia in different media compositions

Table 1: Effect of nursery media on seedling emergence (Days) (%), days to 2-true leaf stage, days to transplanting, seedling height (cm), Leaf area (cm²), Leaf number and LAI of Marigold, Calendula, Dianthus and Salvia

Treatment	Seedling emergence (Days)			Days to 2 leaf stage			Days to transplanting			Seedling height (cm)			Leaf area (cm ²)			Leaf number			LAI		
	2018-19	2019-20	Pooled	2018-19	2019-20	Pooled	2018-19	2019-20	Pooled	2018-19	2019-20	Pooled	2018-19	2019-20	Pooled	2018-19	2019-20	Pooled	2018-19	2019-20	Pooled
Marigold																					
M ₁	4.95	5.28	5.12	14.28	14.31	14.30	33.14	33.81	33.47	4.98	4.32	4.65	2.20	2.84	2.52	3.66	3.66	3.83	0.96	0.96	0.96
M ₂	3.73	4.07	3.90	12.90	12.97	12.93	30.12	30.12	30.12	6.98	6.32	6.65	5.20	6.55	5.88	3.59	3.87	3.79	1.10	1.25	1.18
M ₃	2.99	3.33	3.16	11.45	11.55	11.50	28.48	29.15	28.81	8.98	8.31	8.65	5.80	6.79	6.29	4.10	4.26	4.27	1.20	1.32	1.26
M ₄	4.01	4.01	4.01	13.18	13.25	13.21	29.62	31.62	30.62	5.99	5.32	5.65	4.60	5.01	4.80	4.00	3.78	3.89	1.01	1.03	1.02
S. Ed.±	0.22	0.21	0.20	0.14	0.16	0.15	0.51	0.54	0.48	0.12	0.12	0.12	0.08	0.24	0.11	0.26	0.20	0.21	0.02	0.04	0.02
CD (5%)	0.76	0.73	0.69	0.44	0.49	0.47	1.58	1.66	1.47	0.36	0.36	0.36	0.25	0.75	0.33	0.21	0.33	0.21	0.06	0.14	0.07
Calendula																					
M ₁	5.12	5.25	5.18	14.25	14.45	14.35	32.43	31.76	32.10	5.13	5.47	5.30	3.58	3.60	3.59	4.36	4.40	4.38	1.20	1.18	1.10
M ₂	3.90	4.07	3.98	12.90	13.23	13.07	29.08	28.41	28.75	7.15	7.48	7.32	5.67	5.69	5.68	4.76	4.86	4.81	1.37	1.23	1.26
M ₃	3.09	3.26	3.18	11.49	11.45	11.47	27.99	27.27	27.63	9.31	9.37	9.34	7.11	7.13	7.12	5.27	4.86	5.06	1.38	1.39	1.39
M ₄	4.15	4.35	4.25	13.15	13.45	13.30	30.06	30.02	30.04	6.28	6.61	6.45	6.23	6.26	6.24	4.71	4.71	4.71	1.22	1.24	1.23
S. Ed.±	0.20	0.21	0.20	0.16	0.13	0.14	0.50	0.52	0.50	0.25	0.23	0.24	0.17	0.17	0.17	0.28	0.22	0.24	0.10	0.10	0.10
CD (5%)	0.70	0.72	0.71	0.50	0.41	0.42	1.54	1.61	1.54	0.79	0.70	0.73	0.52	0.51	0.52	0.85	0.67	0.75	NS	NS	NS
Dianthus																					
M ₁	5.62	4.95	5.28	13.07	11.91	12.49	33.47	32.43	32.95	4.32	4.95	4.63	1.87	2.12	1.99	4.00	4.63	4.31	0.86	0.90	0.88
M ₂	4.67	4.07	4.37	11.47	10.50	10.98	29.78	28.78	29.28	6.32	6.91	6.62	4.87	5.08	4.97	6.00	6.63	6.31	1.08	1.12	1.10
M ₃	4.03	3.87	4.10	10.33	9.08	9.71	28.81	27.60	28.21	8.25	8.61	8.43	5.47	5.76	5.61	8.00	8.56	8.28	1.18	1.25	1.22
M ₄	4.71	4.28	4.50	11.57	10.89	11.23	29.95	29.95	29.95	5.29	5.64	5.46	4.27	4.58	4.42	6.00	5.59	5.79	0.89	0.96	0.93
S. Ed.±	0.23	0.34	0.28	0.20	0.26	0.21	0.47	0.66	0.47	0.10	0.39	0.23	0.08	0.21	0.13	0.10	0.47	0.27	0.03	0.04	0.04
CD (5%)	0.79	1.17	0.97	0.61	0.79	0.65	1.44	2.04	1.44	0.31	1.19	0.70	0.25	0.65	0.39	0.31	1.46	0.82	0.11	0.12	0.11
Salvia																					
M ₁	4.62	4.12	4.37	12.50	11.88	12.19	33.48	31.88	32.68	3.00	3.37	3.18	2.10	2.38	2.24	4.21	4.77	4.49	0.34	0.37	0.36
M ₂	3.73	3.04	3.18	9.80	9.53	9.67	30.02	30.18	30.11	5.84	5.99	5.92	3.13	3.72	3.43	4.55	5.32	4.93	0.48	0.51	0.50
M ₃	2.93	3.06	2.84	9.21	9.03	9.12	28.63	27.70	28.16	6.11	6.33	6.22	5.67	6.04	5.85	7.00	6.37	6.95	1.00	1.15	1.08
M ₄	4.20	4.20	4.20	10.58	9.49	10.04	31.73	30.47	31.10	4.45	4.57	4.51	4.96	5.24	5.10	4.36	4.85	4.61	0.62	0.77	0.70
S. Ed.±	0.30	0.50	0.39	0.18	0.52	0.34	0.52	1.27	0.89	0.09	0.13	0.10	0.06	0.34	0.20	0.08	0.45	0.26	0.01	0.06	0.03
CD (5%)	1.04	1.20	1.36	0.55	1.59	1.03	1.60	3.93	2.75	0.27	0.40	0.32	0.20	1.04	0.61	0.26	1.39	0.79	0.03	0.18	0.10

Where, M₁: Cocopeat (60): Vermiculite (20): Perlite (20); M₂: Cocopeat (60): vermicompost (40); M₃: Cocopeat (60): vermicompost (40): Microbial consortium; M₄: Conventional nursery (soil: sand: FYM); S. Ed.: standard error of the differences; CD: Critical difference

Table 2 : Effect of nursery media on root length (cm), Root fresh weight(g), Root dry weight(g), No of roots per seedling, Seedling fresh weight(g), Seedling dry weight(g) and Turgidity (RWC%) of Marigold, Calendula, Dianthus and Salvia

Treatment	Root length (cm)			Root fresh weight(g)			Root dry weight(g)			No of roots per seedling			Seedling fresh weight(g)			Seedling dry weight(g)			Turgidity (RWC%)		
	2018-19	2019-20	Pooled	2018-19	2019-20	Pooled	2018-19	2019-20	Pooled	2018-19	2019-20	Pooled	2018-19	2019-20	Pooled	2018-19	2019-20	Pooled	2018-19	2019-20	Pooled
Marigold																					
M ₁	4.01	4.70	4.36	0.15	0.22	0.18	0.01	0.01	0.01	8.98	9.89	9.44	0.51	0.54	0.53	0.08	0.11	0.10	77.18	78.18	77.68
M ₂	7.02	7.64	7.33	0.30	0.40	0.35	0.09	0.12	0.11	28.98	30.13	29.55	1.49	1.56	1.52	0.36	0.39	0.38	78.61	79.61	79.11
M ₃	9.00	9.78	9.39	0.37	0.47	0.42	0.11	0.15	0.13	37.98	38.47	38.23	1.52	1.62	1.57	0.37	0.44	0.40	79.52	80.52	80.02
M ₄	3.00	4.13	3.57	0.22	0.28	0.25	0.04	0.04	0.04	17.98	18.74	18.36	0.56	0.66	0.61	0.10	0.13	0.12	77.90	78.57	78.23
S. Ed.±	0.11	0.23	0.16	0.01	0.01	0.01	0.01	0.02	0.01	0.45	0.28	0.36	0.02	0.04	0.03	0.01	0.02	0.01	1.32	1.28	1.30
CD (5%)	0.35	0.70	0.50	0.01	0.04	0.02	0.01	0.05	0.03	1.40	0.85	1.12	0.06	0.11	0.08	0.02	0.05	0.03	NS	NS	NS
Calendula																					
M ₁	4.79	4.95	4.87	0.17	0.18	0.18	0.01	0.01	0.01	13.57	14.23	13.90	0.52	0.55	0.54	0.09	0.09	0.09	78.15	78.35	78.25
M ₂	5.73	5.80	5.76	0.26	0.27	0.26	0.10	0.10	0.10	28.69	29.36	29.03	1.49	1.54	1.52	0.38	0.38	0.38	79.04	79.05	79.05
M ₃	8.96	9.13	9.05	0.37	0.38	0.38	0.13	0.14	0.14	33.64	34.31	33.97	1.57	1.57	1.57	0.38	0.38	0.38	80.66	80.29	80.47
M ₄	6.61	6.78	6.70	0.24	0.25	0.24	0.07	0.06	0.06	19.61	20.20	19.91	0.58	0.62	0.60	0.19	0.10	0.14	78.80	78.90	78.85
S. Ed.±	0.30	0.31	0.30	0.01	0.01	0.01	0.01	0.01	0.01	0.59	0.57	0.58	0.02	0.02	0.02	0.03	0.01	0.02	1.51	1.49	1.50
CD (5%)	0.92	0.95	0.94	0.04	0.04	0.04	0.03	0.02	0.03	1.81	1.77	1.79	0.08	0.06	0.06	0.10	0.02	0.06	NS	NS	NS
Dianthus																					
M ₁	8.01	8.31	8.16	0.15	0.16	0.16	0.01	0.01	0.01	9.65	10.23	9.94	0.48	0.54	0.51	0.07	0.08	0.08	76.85	77.68	77.26
M ₂	7.05	7.13	7.26	0.30	0.31	0.31	0.09	0.09	0.09	29.64	30.46	30.05	1.46	1.58	1.52	0.35	0.36	0.36	78.28	79.29	78.78
M ₃	8.87	9.38	9.12	0.37	0.39	0.38	0.11	0.14	0.12	38.65	39.47	39.06	1.49	1.80	1.64	0.36	0.38	0.37	78.85	80.95	79.90
M ₄	6.77	7.11	6.94	0.22	0.23	0.23	0.04	0.04	0.04	18.65	19.74	19.19	0.53	0.64	0.58	0.10	0.09	0.10	77.23	77.90	77.56
S. Ed.±	0.38	0.58	0.47	0.01	0.01	0.01	0.01	0.01	0.01	0.45	0.50	0.47	0.02	0.12	0.06	0.01	0.01	0.01	1.24	1.95	1.58
CD (5%)	1.17	1.78	1.46	0.01	0.03	0.02	0.01	0.03	0.02	1.40	1.54	1.45	0.06	0.38	0.19	0.02	0.03	0.02	NS	NS	NS
Salvia																					
M ₁	5.99	6.44	6.21	0.36	0.39	0.37	0.11	0.12	0.12	23.72	24.57	24.14	0.12	0.13	0.12	0.03	0.05	0.04	77.17	78.02	77.59
M ₂	9.08	9.29	9.18	0.28	0.30	0.29	0.08	0.09	0.08	18.65	20.58	19.61	0.13	0.14	0.14	0.03	0.05	0.04	78.59	79.60	79.09
M ₃	9.36	9.52	9.44	0.45	0.48	0.46	0.14	0.15	0.14	65.17	65.58	65.38	1.28	1.34	1.31	0.33	0.35	0.34	79.35	80.62	79.99
M ₄	4.72	5.08	4.90	0.24	0.27	0.25	0.07	0.08	0.08	16.55	17.48	17.02	0.51	0.57	0.54	0.13	0.13	0.13	77.90	78.56	78.23
S. Ed.±	0.14	0.30	0.21	0.01	0.02	0.01	0.00	0.01	0.00	0.54	1.25	0.86	0.01	0.03	0.02	0.002	0.01	0.01	1.30	1.88	1.59
CD (5%)	0.42	0.92	0.66	0.02	0.06	0.04	0.01	0.02	0.01	1.67	3.85	2.65	0.03	0.09	0.06	0.007	0.03	0.02	NS	NS	NS

Where, M₁: Cocopeat (60): Vermiculite (20): Perlite (20); M₂: Cocopeat (60): vermicompost (40); M₃: Cocopeat (60): vermicompost (40): Microbial consortium; M₄: Conventional nursery (soil: sand: FYM); S. Ed.: standard error of the differences; CD: Critical difference

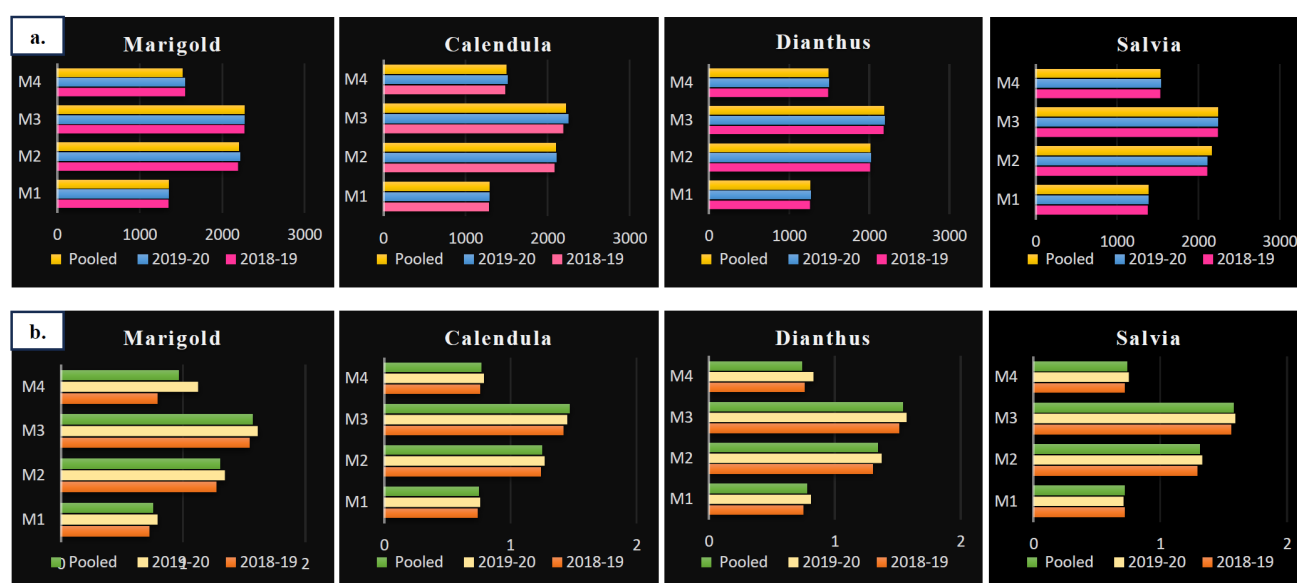


Fig 2.: a. Seedling vigour index and b. Chlorophyll content (mg g^{-1} FW) of Marigold, Calendula, Dianthus and Salvia

Table 3 : Physico-Chemical characteristics of the media used in the experiment

Treatments	pH		EC (dS/m)		CEC (m eq/100g)		Porosity (%)		Water Holding Capacity %	
	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019
M ₁ – Cocopeat (60): Vermiculite (20): Perlite (20)	6.59	6.43	0.049	0.048	17.48	17.06	61.66	60.16	62.27	60.75
M ₂ – Cocopeat (60): vermicompost (40)	5.82	5.68	0.041	0.038	8.83	8.61	56.47	55.09	54.09	52.77
M ₃ – Cocopeat (60): vermicompost (40): Microbial consortium	5.59	5.45	0.031	0.034	13.21	12.89	58.62	57.20	52.72	51.44
M ₄ – Conventional nursery (soil: sand: FYM)	5.31	5.32	0.023	0.025	4.56	4.44	47.16	46.02	37.4	35.9

Discussion

Growing media affects the germination of seeds and the subsequent emergence and development of seedlings in a nursery (Bhardwaj, 2014), as it serves as a reservoir for moisture and essential plant nutrients (Kazemi and Mohorko, 2017). All seeds require water, oxygen, and appropriate temperature conditions for successful germination. Water is essential for softening the protective seed coat, while oxygen is needed for aerobic respiration to generate energy for both germination and growth. The embryo derives energy by metabolizing its stored food reserves.

In the context of Marigold, Calendula, Dianthus, and Salvia, the M₃ growing media demonstrated excellent water retention capabilities, providing a consistent moisture supply alongside adequate porosity. This allowed for proper moisture retention and gas exchange between the media and the seeds, thus facilitating improved seedling emergence. Comparable findings were reported by Hota and Arulmozhiselvan (2017) concerning tomato

plants. Subsequently, true leaves appear shortly after the cotyledons and begin to produce energy through photosynthesis, which nourishes the plant and assists in reaching the transplantable stage promptly, alongside improved physical properties and enhanced levels of nutrients, water, and air in the seedling root zone of M₃ (Bharadwaj, 2013b). The sufficient aeration provided by a combination of cocopeat and vermicompost, along with the substantial nutrient availability enhanced by vermicompost and microbial consortia, including *Azotobacter*, *Azospirillum*, and PSB in M₃ media, resulted in taller plants with enhanced leaf production (Nissi, 2018). This increase in leaf surface area ultimately led to a higher Leaf Area Index (Patel *et al.*, 2019). The heightened leaf production of seedlings may correlate with a simultaneous increase in plant height (Moles *et al.*, 2009). Additionally, the microorganisms present in the vermicompost and microbial consortia within the M₃ nursery media synthesize plant growth hormones, primarily auxins, gibberellins, and cytokinins. The maximum growth of both roots and shoots may be attributed to the higher

availability of auxin in this growing medium, derived from the consortium as well as from the humic acid and humates found in vermicompost (Scaglia *et al.*, 2016). Furthermore, the intensive activity of earthworms in vermicompost results in nutrient levels such as nitrogen, phosphorus, and potassium being 5 to 11 times greater than those in ordinary soil. Kiyasudeen *et al.*, 2016 also noted that the application of vermicompost enhances both micro and macronutrient content of the media, thereby potentially improving shoot growth, as reflected in the dry and fresh weight of the shoots. Additionally, the incorporation of cocopeat improved the physical and chemical properties of the media by reducing its density and increasing porosity, which encouraged the formation of a greater number of roots with enhanced dry and fresh weight. These findings align with earlier research conducted by Bachman and Metzger (2008), Gupta *et al.* (2023), and Kispotta (2019) on Marigold and cornflower; Tomato; and African and French Marigold. Similarly, Baskaran *et al.* (2004) reported comparable results when utilizing a media combination of coir compost and vermicompost (1:1) for chrysanthemum cuttings. The higher vigor index observed in M₃ is likely attributable to improved germination and seedling height, contributing to an enhanced vigor index. The outcomes of this research are consistent with the findings of Prajapati *et al.* (2017) and Parasana *et al.* (2013).

Conclusion

It could be concluded that the healthy status, and quality nursery seedling production of Marigold (Inca Orange), Calendula (Bon Bon Mix), Dianthus (Ideal Select Mix) and Salvia (Vista Mix) could be dramatically influenced by the component of the growing media. Findings of the current research exhibited that the media substrate of Cocopeat and vermicompost in the ratio of 60:40 along with Microbial consortium (M₃) was particularly suitable for sturdy and healthy nursery seedling production for all four annuals.

Author contribution: Data collection and manuscript preparation contributed by D. Sarkar,

R. Borbaruah, P. Hatibarua, edited by M. C. Talukdar, D. Naikwad, D. J. Nath

Conflict of interest: The authors declare that they have no conflict of interest.

References

- Abdul Baki, A.A. & Anderson, J.D. (1973). Vigour determination in soybean seed by multiple criteria. *Crop. Sci.* **13**, 630-633.
- Bachman, G.R. & Metzger, J.D. (2008). Growth of bedding plants in commercial potting substrate amended with vermicompost. *Biores. Tech.* **99**, 3155-3166.
- Baskaran, V.; Janakiram, T. and Jayanthi, R. (2004). Correlation and path analysis studies in chrysanthemum. *J. Orn. Hort.*, **7**(3-4), 37-44.
- Bharadwaj, R.L. (2013b). Effect of nine different propagation media on seed germination and the initial performance of papaya (*Carica papaya* L.) seedlings. *J. Hort. Sci. Biotech.* **88**(5), 531-536.
- Bhardwaj, R.L. (2014). Effect of growing media on seed germination and seedling growth of papaya cv. 'Red lady'. *African journal of plant science*, **8**(4), 178-184.
- Black, C.A. (ed.) (1965). Method of Soil Analysis, Part 2, Chemical and Microbiological Properties, American Society of Agronomy, Inc, Publisher, Madison, Wisconsin USA.
- Evans, S. (1972). General geomorphometry, derivatives of altitude, and descriptive statistics in Chorley, R.J., Ed., spatial analysis in geomorphology, Methuen & Co. Ltd., London, pp. 17-90.
- Gupta, A., Qureshi, N., Dangwal, A., Kumar, C., & Badoni, A. (2023). Nutrient analysis of different media used for nursery production. *International Journal of Plant & Soil Science*, **35**, 728-734.
- Hota, S. and Arulmozhiselvan, K. (2017). Standardization of soilless growth media under capillary rise irrigation principle for nursery raising of tomato (*Lycopersicon esculentum*) seedlings by nurseries pack. *Curr. Adv. Agri. Sci.* **9**(1), 101-103.
- Jankowska, B. (2010). Effect of intercropping white cabbage with french marigold (*Tagetes patula* nana) and pot marigold (*Calendula officinalis*) on diamondback moth (*Plutella xylostella* L.) population density and its parasitoid complex. *Vegetable Crops Research Bulletin*, **73**, 107.
- Kalaiselvam I, Devaraj A. 2011. Effect of root exudates of *Tagetes* sp. on egg hatching behaviour of *Meloidogyne incognita*. *Int Res J Pharm.* **2**, 93-96.
- Kazemi, F., & Mohorko, R. (2017). Review on the roles and effects of growing media on plant performance in green roofs in world climates. *Urban Forestry & Urban Greening*, **23**, 13-26.
- Kispotta, G.S. (2019). Effect of different rooting media on propagation and performance of African marigold (*Tagetes erecta* L.) cv. Pusa Basanti Gaiinda through stem cutting. Ph.D. Thesis. Indira Gandhi Krishi Vishwavidyalaya, Raipur (CG).
- Kiyasudeen S. K., Ibrahim, M. H., Quaik, S., Ahmed Ismail, S., S. K. K., Ibrahim, M. H., ... & Ismail, S. A. (2016). Vermicompost, its applications and derivatives. *Prospects*

- of organic waste management and the significance of earthworms, 201-230.
- Mahmoud, T. S. M., Nabila, E. K., Rayya, M. A., & Eisa, R. A. (2019). Effect of planting dates and different growing media on seed germination and growth of pistachio seedlings. *Bulletin of the National Research Centre*, **43**, 1-6.
- Abdul Baki, A.A. & Anderson, J.D. (1973). Vigour determination in soybean seed by multiple criteria. *Crop Science*, **13**, 630-633.
- Bachman, G.R. & Metzger, J.D. (2008). Growth of bedding plants in commercial potting substrate amended with vermicompost. *Bioresource Technology*, **99**, 3155-3166.
- Baskaran, V., Janakiram, T. & Jayanthi, R. (2004). Correlation and path analysis studies in chrysanthemum. *Journal of Ornamental Horticulture*, **7**(3-4), 37-44.
- Bharadwaj, R.L. (2013b). Effect of nine different propagation media on seed germination and the initial performance of papaya (*Carica papaya* L.) seedlings. *The Journal of Horticultural Science and Biotechnology*, **88**(5), 531-536.
- Bhardwaj, R.L. (2014). Effect of growing media on seed germination and seedling growth of papaya cv. 'Red lady'. *African journal of plant science*, **8**(4), 178-184.
- Black, C.A. (ed.) (1965). Method of Soil Analysis, Part 2, Chemical and Microbiological Properties, American Society of Agronomy, Inc, Publisher, Madison, Wisconsin USA.
- Evans, S. (1972). General geomorphometry, derivatives of altitude, and descriptive statistics in Chorley, R.J., Ed., spatial analysis in geomorphology, Methuen & Co. Ltd., London, pp. 17-90.
- Gupta, A., Qureshi, N., Dangwal, A., Kumar, C., & Badoni, A. (2023). Nutrient analysis of different media used for nursery production. *International Journal of Plant & Soil Science*, **35**, 728-734.
- Hota, S. & Arulmozhiselvan, K. (2017). Standardization of soilless growth media under capillary rise irrigation principle for nursery raising of tomato (*Lycopersicon esculentum*) seedlings by nutrised pack. *Current Advances in Agricultural Sciences*, **9**(1), 101-103.
- Jankowska, B. (2010). Effect of intercropping white cabbage with french marigold (*Tagetes patula* nana) and pot marigold (*Calendula officinalis*) on diamondback moth (*Plutella xylostella* L.) population density and its parasitoid complex. *Vegetable Crops Research Bulletin*, **73**, 107.
- Kalaiselvam I, Devaraj A. (2011). Effect of root exudates of *Tagetes* sp. on egg hatching behaviour of *Meloidogyne incognita*. *International Research Journal of Pharmacy*, **2**, 93-96.
- Kazemi, F., & Mohorko, R. (2017). Review on the roles and effects of growing media on plant performance in green roofs in world climates. *Urban Forestry & Urban Greening*, **23**, 13-26.
- Kispotta, G.S. (2019). Effect of different rooting media on propagation and performance of African marigold (*Tagetes erecta* L.) cv. Pusa Basanti Gaiinda through stem cutting. Ph.D. Thesis. Indira Gandhi Krishi Vishwavidyalaya, Raipur (CG).
- Kiyasudeen S, K., Ibrahim, M. H., Quaik, S., Ahmed Ismail, S., S, K. K., Ibrahim, M. H., ... & Ismail, S. A. (2016). Vermicompost, its applications and derivatives. Prospects of organic waste management and the significance of earthworms, 201-230.
- Mahmoud, T. S. M., Nabila, E. K., Rayya, M. A., & Eisa, R. A. (2019). Effect of planting dates and different growing media on seed germination and growth of pistachio seedlings. *Bulletin of the National Research Centre*, **43**, 1-6.
- Manzoor, S., Rashid, R., Panda, B. P., Sharma, V., & Azhar, M. (2022). Green extraction of lutein from marigold flower petals, process optimization and its potential to improve the oxidative stability of sunflower oil. *Ultrasonics sonochemistry*, **85**, 105994.
- Mathowa, T., Tshagofatso, N., Mojeremane, W., Matswane, C., Legwaila, G. M., & Oagile, O. (2016). Effect of commercial growing media on emergence, growth and development of tomato seedlings. *International Journal of Agronomy and Agricultural Research*, **9**(1), 83-91.
- Miraj, S., & Kiani, S. (2016). A review study of therapeutic effects of *Salvia officinalis* L. *Der Pharmacia Lettre*, **8**(6), 299-303.
- Moles, A. T., Warton, D. I., Warman, L., Swenson, N. G., Laffan, S. W., Zanne, A. E., ... & Leishman, M. R. (2009). Global patterns in plant height. *Journal of ecology*, **97**(5), 923-932.
- Nath, J. C., Deka, K. K., Maheswarappa, H. P., & Sumitha, S. (2019). System productivity enhancement in coconut (*Cocos nucifera*) garden by intercropping with flower crops in Assam. *Indian Journal of Agricultural Sciences*, **89** (11), 1842-5.
- Nissi, F.G. (2018). Growth of tomato (*Solanum lycopersicum* L.) seedlings in different potting mixes, under hi-tech nursery in green house conditions. *International Journal of Pure & Applied Bioscience*, **6**(5), 692-695.
- Parasana, J. S., Leua, H. N., & Ray, N. R. (2013). Effect of different growing medias mixture on the germination and seedling growth of mango (*Mangifera indica*) cultivars under net house conditions. *The Bioscan*, **8**(3), 897-900.
- Patel, M.V.; Parmar, B.R.; Halpati, A.P.; Parmar, A.B. and Pandey, A.K. (2019). Effect of growing media and foliar spray of organics on seedling growth and vigour of acid lime. *International Journal of Chemical Studies*, **7**(1), 01-04.
- Pires, T. C., Dias, M. I., Barros, L., Calhella, R. C., Alves, M. J., Oliveira, M. B. P., ... & Ferreira, I. C. (2018). Edible flowers as sources of phenolic compounds with bioactive potential. *Food Research International*, **105**, 580-588.
- Prajapati, D.G.; Satodiya, B.N.; Desai, A.B. and Nagar, P.K. (2017). Influence of storage period and growing media on seed germination and growth of acid lime seedlings (*Citrus aurantifolia* Swingle) cv. Kagzi. *Journal of Pharmacognosy and Phytochemistry*, **6**(4), 1641-1645.
- Rasool, S., Ahmad, I., Ziaf, K., & Naveed, M. (2024). Plant growth promoting rhizobacteria incorporated soilless substrates—A potential arena for high quality nursery production of ornamentals. *Scientia Horticulturae*, **326**, 112745.
- Scaglia, B.; Nunes, R.R. and Rezende, M.O.O. (2016). Investigating organic molecules responsible of auxin-like activity of humic acid fraction extracted from vermicompost. *Science of the Total Environment*, **562**, 289-295.
- Sharma, S., Das, A., Kumari, A., & Gupta, M. M. (2024). Technological insights into lutein isolation from marigold

- flower and their diverse applications, a compendious review. *Phytochemistry Reviews*, 1-22.
- Singh, K.P. and Singh, R.P. (1977). The chlorophyll content of sun and shade leaves of common tree growing at Varanasi, India. *Indian Journal of Ecology*, **4**, 46-54.
- Thakur, S., Pathania, S., & Kumar, A. Floral Culture and Cut Flower Production. *New Horizons and Advancements in Horticulture Volume*, 320. (2023)
- Ullah, M. A., Hassan, A., & Hamza, A. (2023). Calendula (*Calendula officinalis*) marigold as medicinal plant. *Journal of Orthopaedic Case Reports*, **2**(9).
- Walia, S., & Kumar, R. (2020). Wild marigold (*Tagetes minuta* L.) an important industrial aromatic crop, liquid gold from the Himalaya. *Journal of Essential Oil Research*, **32**(5), 373-393.
- Xing, J., Liu, C., Zhang, Y., He, H., Zhou, Y., Li, L., ... & Xiang, W. (2015). *Sphaerisporangium dianthi* sp. nov., an endophytic actinomycete isolated from a root of *Dianthus chinensis* L. *Antonie van Leeuwenhoek*, **107**, 9-14.